

## SYNTHESES OF TWO ENANTIOMERIC PAIRS OF MYO-INOSITOL(1,2,4,5,6) AND -(1,2,3,4,5) PENTAKISPHOSPHATE

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Abstract: Two enantiomeric pairs of myo-inositol(1,2,4,5,6)P<sub>5</sub> and -(1,2,3,4,5)P<sub>5</sub> have efficiently been synthesized by means of the lipase catalyzed acetylation of 1,2.5,6-di-O-isopropylidene-myo-inositol and the benzoyl migration procedure. © 1998 Elsevier Science Ltd. All rights reserved.

Since the discovery that D-myo-inositol-1,4,5-trisphosphate [Ins(1,4,5)P<sub>3</sub>] plays a pivotal role as a second messenger in the transmembrane signaling, thus mobilizing calcium ions from the intracellular storage, its interaction with the I(1,4,5)P<sub>3</sub> receptor and metabolic enzymes has been a subject of intensive investigations. One of the major metabolic pathways involves a specific phosphorylation of Ins(1,4,5)P3 to Ins(1,3,4,5)P<sub>4</sub> by Ins(1,4,5)P<sub>3</sub>-3-kinase. Although IP<sub>5</sub>s were not recognized as naturally occurring metabolites of IP<sub>3</sub> and IP<sub>4</sub> until recently, their biological roles and functional importances have been implicated in many biological systems.3 In addition, some of the synthetic IP5 regioisomers such as D/L-Ins(1,2,3,4,5)P<sub>5</sub> (2) were found to show high affinities toward the D-Ins(1,3,4,5)P<sub>4</sub> receptor protein purified from pig cerebellum. There exist six possible IP<sub>5</sub> regioisomers: two meso compounds [Ins(1,3,4,5,6)P<sub>5</sub>, Ins(1,2,3,4,6)P<sub>5</sub>] and two pairs of enantiomers [D/L-Ins(1,2,4,5,6)P<sub>5</sub>, D/L-Ins((1,2,3,4,5)P<sub>5</sub>]. Several groups have reported syntheses of meso and racemic IP5 isomers,5 including the synthesis of all possible regioisomers of IP5 based on the benzoyl group migration method.<sup>6</sup> Very recently, the first synthesis of chiral IP<sub>5</sub>s via the camphanate ester resolution route was reported.<sup>7</sup> We wish to report herein our efforts on the synthesis of the two enantiomeric pairs of Ins(1,2,4,5,6)P<sub>5</sub> (1) and Ins(1,2,3,4,5)P<sub>5</sub> (2)

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Our synthetic approaches to homochiral 1 and 2 are based on the enzyme catalyzed asymmetric acetylation of 1,2:5,6-di-O-isopropylidene-myo-inositol (3). Thus, racemic diol 3<sup>8</sup> in diethyl ether was subjected to acetic anhydride in the presence of lipase from Candida rugosa (Sigma, CRL). The reaction was stopped at ca. 50% completion, and the product was filtered through celite and chromatographed on silica gel to give the unreacted diol (4D, 46%, 87% ee) and the monoacetylated product (5L, 48%, 84% ee). Hydrolysis of 5L with LiOH in aqueous methanol gave 4L in good yield. The optical purities of 4D and 4L could be improved to 98% ee upon recrystallization from hexane and CHCl<sub>3</sub> (1:1) in ca. 70% recovery. The absolute configurations of 4D and 4L were determined on the basis of the HPLC retention time on a Chiralcel OD column, after their conversion to the I(1,4,5,6)Bz<sub>4</sub> derivatives. Thus, benzoylation of 4D with excess BzCl in pyridine, followed by a) acid-catalyzed partial solvolysis (p-TsOH, MeOH-CH<sub>2</sub>Cl<sub>2</sub>) of the trans-acetal of 4D-Bz<sub>2</sub>, b) further benzoylation, and c) acid-catalyzed removal of the cis-acetal gave D-I(1,4,5,6)Bz<sub>4</sub>. Similarly, 4L was converted to L-I(1,4,5,6)Bz<sub>4</sub> was 9.82 (Chiralcel OD column, iPrOH-heptane 1:3, flow rate 2.0 ml/min), in accord with the reported order of retention times. The intention times of particles are provided to the reported order of retention times.

Scheme 1. a. CRL, Ac<sub>2</sub>O/Et<sub>2</sub>O, RT. b. LiOH, H<sub>2</sub>O-MeOH, 0 °C.

Chiral diol **4D** was monobenzoylated under the conventional conditions employing BzCl in pyridine to give a mixture of **6D** and **7D** (in 91:9 ratio based on <sup>1</sup>H-NMR, 82% yield). The base-catalyzed benzoyl migration<sup>11</sup> of the crude product shifted the ratio to **6D**: **7D** = 64:36.<sup>12</sup> After column chromatography, **6D** and **7D** each was hydrolyzed in hot aqueous acetic acid, and the product was phosphorylated by successive reactions with diethyl chlorophosphite and diisopropylethylamine in DMF, and then 30% H<sub>2</sub>O<sub>2</sub> to afford **8L** and **9D**.<sup>13</sup> In the final step, all protecting groups were removed by successive treatments with TMSBr and then LiOH. The sodium salt of the target compounds **1L** and **2D** were obtained after ion exchange chromatography on Dowex 50x8-100 (H<sup>+</sup> form), pH adjustment to 10 with NaOH, and lyophilization (Scheme 2).<sup>14</sup> Compound **4L** was analogously transformed to **1D** and **2L**.<sup>14</sup>

Scheme 2. a. BzCl, pyridine, 82% (sum of **6D** and **7D**). b. pyridine-H<sub>2</sub>O (6:4), 100 °C, 1h. c. (i) 80% aq. AcOH, 100 °C, 1h. (ii) (EtO)<sub>2</sub>P-Cl, iPr<sub>2</sub>NEt, DMF. (iii) H<sub>2</sub>O<sub>2</sub>, ~50%. d. (i) TMSBr, CH<sub>2</sub>Cl<sub>2</sub>. (ii) 1N LiOH, 80 °C, 3h. (iii) H<sup>+</sup> ion-exchange. (iv) NaOH, pH 10, quant.

In sum, we have successfully prepared each enantiomer of I(1,2,4,5,6)P<sub>5</sub> (1D and 1L) and I(1,2,3,4,5)P<sub>5</sub> (2D and 2L) via the CRL catalyzed asymmetric acetylation of 1,2:5,6-di-O-isopropylidene-myo-inositol and the benzoyl migration procedure.

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## References and Notes

\* Dedicated to Professor Robert M. Coates (University of Illinois) on the occasion of his 60th Birthday.

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- 9. The CRL catalyzed acetylation could routinely be run in 5-10 g scales. 4D: mp 151-153 °C, [α]<sub>D</sub><sup>27</sup> +9.12 (c 0.74, CHCl<sub>3</sub>); 4L: mp 151-153 °C, [α]<sub>D</sub><sup>28</sup> -8.85 (c 1.0, CHCl<sub>3</sub>). A similar but smaller scale resolution of 1,2:5,6-dicyclohexylidene-myo-inositol with bovine pancreas cholesterol esterase was previously reported: Liu, Y.-C.; Chen, C.-S. Tetrahedron Lett. 1989, 30, 1617-1620.
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- 12.  $R_f$  values for **6D** and **7D** are 0.2 and 0.25 (ethyl acetate : n-hexane = 1 : 2). **6D**: mp 183-186 °C,  $[\alpha]_D^{27}$  -24.4 (c 0.53, CH<sub>3</sub>OH); **7D**: mp 139-142 °C,  $[\alpha]_D^{27}$  +6.9 (c 0.62, CH<sub>3</sub>OH); **6L**: mp 184-186 °C,  $[\alpha]_D^{27}$  +23.8 (c 0.63, CH<sub>3</sub>OH); **7L**: mp 142-143 °C,  $[\alpha]_D^{27}$  -6.3 (c 0.69, CH<sub>3</sub>OH).
- 13. **8L**:  $[\alpha]_D^{27}$  -12.5 (c 0.62, CH<sub>2</sub>Cl<sub>2</sub>); **9D**:  $[\alpha]_D^{27}$  +6.0 (c 0.60, CH<sub>2</sub>Cl<sub>2</sub>); **8D**:  $[\alpha]_D^{27}$  +13.7 (c 1.58, CH<sub>2</sub>Cl<sub>2</sub>); **9L**:  $[\alpha]_D^{27}$  4.6 (c 1.43, CH<sub>2</sub>Cl<sub>2</sub>).
- 14. **1D**:  $[\alpha]_D^{25}$  -6.0 (c 0.40, H<sub>2</sub>O, pH 10), lit.  $[\alpha]_D^{24}$  -7.1 (c 0.83, H<sub>2</sub>O, pH 1.6)<sup>7</sup>; **1L**:  $[\alpha]_D^{25}$  +7.5 (c 0.40, H<sub>2</sub>O, pH 9.5), lit.  $[\alpha]_D^{24}$  -6.2 (c 0.96, H<sub>2</sub>O, pH 1.6)<sup>7</sup>; **2D**:  $[\alpha]_D^{25}$  -5.0 (c 0.40, H<sub>2</sub>O, pH 9.2), lit.  $[\alpha]_D^{24}$  -4.0 (c 0.23, H<sub>2</sub>O, pH 1.6)<sup>7</sup>; **2L**:  $[\alpha]_D^{25}$  +5.8 (c 0.40, H<sub>2</sub>O, pH 9.5), lit.  $[\alpha]_D^{24}$  +4.3 (c 0.43, H<sub>2</sub>O, pH 1.6)<sup>7</sup>.